Enhancement of solubility and dissolution rate of poorly water-soluble domperidone by the formulation of...
Enhancement of Solubility and Dissolution Rate of Poorly Water-Soluble Domperidone by the Formulation of Multicomponent Solid Dispersion Using Solvent Evaporation Method


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ABSTRACT

First-pass metabolism affects many oral medications and limits the attainment of their therapeutic level. It can be bypassed by administering buccal dosage forms that allow systemic drug absorption via buccal mucosa. Drugs formulated as buccal medicaments should have an acceptable solubility in saliva. Numerous technologies had been experimented to increase the aqueous solubility of poorly water-soluble drugs e.g. solid dispersion technique. This technique is efficient for improving the solubility and dissolution rate of hydrophobic drugs and consequently improving their bioavailability. Domperidone is an antiemetic drug that undergoes extensive first-pass metabolism, having poor solubility in saliva and poor bioavailability. This study aimed to improve the aqueous solubility of domperidone at pH simulating saliva by preparing multicomponent solid dispersions using different carriers by solvent evaporation method. In-vitro dissolution studies showed enhanced dissolution rates of all prepared systems with release kinetics approaching Higuchi model. Ternary solid dispersion (SD) of 1:9:0.25 drug/polyvinylpyrrolidone K30/pluronic F-127, respectively, achieved the highest dissolution rate. Physicochemical characterization of this SD using differential scanning calorimetry, Fourier-transform infrared spectroscopy, powder X-ray diffraction and scanning electron microscopy indicated the presence of an interaction between domperidone and polyvinylpyrrolidone K30 with evidence of drug amorphization that might be responsible for the enhanced dissolution rate.

Keywords: Domperidone, pluronic F-127, solvent evaporation method, multicomponent solid dispersions, physicochemical characterization.

INTRODUCTION

First-pass metabolism is the most popular disadvantage of the orally administrated drugs where this pathway affects drug bioavailability 1. Alternative non-enteral routes of administration can overcome this metabolic pathway allowing the systemic drug absorption, thereby increasing its bioavailability and decreasing metabolite production e.g. sublingual, rectal, inhalation, intravenous, intramuscular and transdermal routes 2.

For a drug to be absorbed, it should have an acceptable solubility at the absorption site. Since many drugs discovered by the technological innovation of combinatorial chemistry are poorly water-soluble entities, it is often difficult to adopt them as candidates for pharmaceutical preparations 2. Therefore, several techniques were developed to improve the aqueous solubility of these drugs. The most popular approaches are the incorporation of the active hydrophobic component into solid dispersions 3, inclusion complexes 4, inert lipid vehicles 5, surfactant dispersion 6, self-emulsifying formulations 7, dry emulsions 8 and niosomes 9.

Solid dispersion systems were defined as the dispersion of one or more active ingredients in an inert carrier matrix at solid state 10. Solid dispersions can be prepared by different methods using different water-soluble carriers. These solid systems exhibit enhanced solubility and dissolution rate compared to the plain drug that may be attributed to the molecular/colloidal dispersion of drug in mixture, absence of aggregation of drug particles, particle size reduction, improved wettability and dispersability and polymorphic transformation of drug crystals 11-13. Enhancement of solubility may contribute directly to the improved bioavailability of poorly water-soluble drugs.

Domperidone (DMP), the model drug of this research, is an antiemetic drug that has the chemical structure of 5-Chloro-1-[1-[3-(2-oxobenzimidazolin-1-yl)propyl]-4-piperidyl] benzimidazolin-2-one (Figure 1). It is described as a peripheral antidopaminergic drug that is mainly used as an antiemetic for the treatment of nausea and vomiting of various etiologies. DMP has low systemic bioavailability about 13-17% of the orally administrated dose due to the extensive hepatic and intestinal metabolism 14.

![Figure 1: Chemical structure of domperidone](image)

Different attempts were performed to improve DMP solubility and hence its bioavailability. For example, the...
solubility enhancement of domperidone was examined using different carriers e.g. polyethylene glycol 4000, polyethylene glycol 6000 and Myrj 52 by melt granulation technique. In addition, multicomponent inclusion complexes of DMP were prepared using native cyclodextrin, cyclodextrin derivatives, hydroxypropyl cellulose, citric acid and other polymers by kneading method resulting in almost 92-100% of domperidone released after 5-60 minutes. The objective of the present study was to improve the solubility and dissolution rate of DMP in phosphate buffer of pH 6.8 by the formulation of solid dispersions (SDs). This pH was selected to simulate salivary pH that ranges from 5.5 to 7.0 in order to incorporate the optimized solid dispersion later into buccal dosage forms. These SDs were prepared by solvent evaporation method using different water-soluble carriers in different weight ratios. In-vitro dissolution studies were performed to select the best formula that in turn would be physicochemically characterized by differential scanning calorimetry (DSC), Fourier-transform infrared spectroscopy (FTIR), powder X-ray diffraction (PXRD) and scanning electron microscopy (SEM). To survey more precisely the mechanism of drug release from the optimized SDs, their in-vitro dissolution data were fitted to zero order, first order and Higuchi kinetic model.

MATERIALS AND METHODS

Materials
Domperidone was given as a gift from Delta Pharma Company for Pharmaceutical Industries, Cairo, Egypt. Dichloromethane was purchased from Fisher Scientific, LTD, Leicestershire, UK. Polyvinylpyrrolidone K30 was supplied by Himedia laboratories PVT, LTD, Mumbai, India. Methanol AR, monobasic potassium hydrogen phosphate, sodium hydroxide pellets and urea were obtained from EL Gomhouria Co., Cairo, Egypt. Anhydrous calcium chloride and pluronic-F-127 were supplied by sigma-aldrich Inc., Missouri, USA. Polyethylene glycol 8000 was purchased from Scharlau Chemie, S.A., Barcelona, Spain. Hydroxypropyl methylcellulose E50 LV was supplied by LOBA Chemie PVT, LTD, Mumbai, India. All other ingredients were of analytical grade.

Phase solubility studies
An excess amount of DMP was added to 20 ml carrier solutions ranging in concentration from 1% to 5% w/v prepared in phosphate buffer solution that was adjusted at pH 6.8 using 0.2 M sodium hydroxide solution in a series of 50 ml stoppered glass bottles. The prepared suspensions were shaken at 25±0.5° C for 7 days in Julabo thermostatically controlled shaking water bath (Julabo SW 20C, Osaka, Japan). After equilibrium being achieved, aliquots were withdrawn, filtered through 0.45 µm syringe filters (0.45 PTFE, Thermo Scientific Chromacol, Leicestershire, UK) and assayed spectrophotometrically at wavelength of 284 nm using Shimadzu UV/VIS spectrophotometer (UV- 1650 PC, Shimadzu Corporation, Kyoto, Japan). DMP content was determined using the regression equation of the standard curve that was developed in the same medium. Blank solutions were performed in the same concentrations of the respective carriers in pH 6.8 phosphate buffer solution. In addition, the solubility of DMP alone was also determined by the same procedure mentioned above.

To investigate the effect of the auxiliary substances e.g. PL F-127, HPMC E50 LV and PEG 8000 on DMP solubility, the previously mentioned solubility phase study was performed using phosphate buffer solution containing 5% w/v PVP K30 and increasing consecration of PL F-127 (ranging from 2% to 4.5% w/v), HPMC E50 LV and PEG 8000 (ranging from 0.5% to 2% w/v).

Preparation of solid dispersions (SDs) by solvent evaporation method
To prepare SDs of DMP with PEG 8000, urea and PVP K30 in weight ratios of 1:1, 1:5 and 1:9; an appropriate amount of carrier was added to a solution of DMP in methanol and dichloromethane (1:1 v/v). This solution was stirred on a magnetic stirrer (1200, Jenway, Staffordshire, UK) for 2 hours at room temperature and maintained in an open tray for at least 12 hours to allow slow evaporation of solvent. After drying overnight, solid residue was scratched, dried in a vacuum oven for 24 hours at room temperature, pulverized and sieved using Tongxin 45-mesh sieve (TX Tongxin, Henan, China).

Powdered samples were stored in closed containers away from the light and humidity and kept in a desiccator containing anhydrous calcium chloride as a dehydrating agent until further evaluation. SDs containing DMP, PVP K30 and PL F-127 in weight ratios of 1:9:0.125, 1:9:0.25 and 1:9:0.5 were prepared as mentioned before. SDs containing DMP, PVP K30, HPMC E50 LV or PEG 8000 in weight ratios of 1:9:2.25, 1:9:4.5 and 1:9:9 were similarly prepared.

Preparation of physical mixtures (PMs)
PMs were prepared by simple trituration of the drug and carriers with their respective weight ratios in a porcelain mortar for 5 minutes. PMs were sieved and stored as mentioned before until use.

Determination of drug content uniformity of the prepared systems:
Powdered samples equivalent to 10 mg of DMP were accurately weighed, dissolved in 50 ml of phosphate buffer (pH 6.8) and stirred on a magnetic stirrer for 15 minutes. These solutions were filtered through 0.45 µm syringe filters, diluted and assayed spectrophotometrically at wavelength of 284 nm for DMP content.

In-vitro dissolution studies
In-vitro dissolution studies of plain DMP, SDs and PMs were performed using dissolution USP apparatus II (rotating paddle) (SOTAX AT7 smart, Allschwil,
The dissolution medium consisted of 500 ml of phosphate buffer (pH 6.8). The stirring speed was 100 rpm and temperature was maintained at 37±0.5°C. Powdered samples of each preparation equivalent to 10 mg of DMP were sprinkled on the surface of the dissolution medium. At the appropriate time intervals for a period of 60 minutes, 3 ml aliquots were withdrawn from the dissolution medium through 0.45 µm syringe filters and replaced with an equivalent amount of fresh medium to keep the volume constant. Concentrations of DMP were determined spectrophotometrically at wavelength of 284 nm. Each experiment was carried out in triplicates to determine the mean and the standard deviation.

The dissolution profiles were evaluated according to four parameters: i) initial dissolution rate (IDR) that was calculated as the percentage of drug dissolved over the first 15 minutes per minute; ii) percentage of drug dissolved after 2 minutes (PD_2); iii) percentage of drug dissolved after 10 minutes (PD_10) and iv) dissolution efficiency (DE_{60%}) parameter after sixty minutes.

Fourier-transform infrared spectroscopy (FTIR)

FTIR spectra of the pure drug, optimized ternary SD, its PM and their individual components were obtained using JASCO FTIR spectrophotometer (FTIR 4100, JASCO, Essex, UK) operated with potassium bromide disc technique. FTIR analysis was performed using a pressure of 6-8 tons, die size of 13 mm, scanning range of 400-4000 cm⁻¹ and resolution of 1 cm⁻¹.

Differential scanning calorimetry (DSC)

DSC analysis was performed using Shimadzu differential scanning calorimeter (DSC-50, Shimadzu Corporation, Kyoto, Japan). Samples (1.5-2.5 mg) were heated in a hermetically sealed aluminum pans at a temperature ranged from 30°C to 300°C and constant rate of 10°C C/min under a nitrogen purge (30 ml/min.).

Powder X-ray diffraction (PXRD):

PXRD patterns were obtained using XGEN X-ray powder diffractometer (XGEN 4000, Scintage Inc., California, USA) supplied with CuKa radiation. Diffractograms were run at a scanning rate of 1.8 degree min⁻¹ and the scanning scope was over a range of 2θ angle from 0 to 80° at room temperature.

A relationship was established between some representative peak heights in the diffraction patterns of the ternary systems and those of a reference substance (i.e., plain drug). This relationship was translated into the following equation that calculates the relative degree of crystallinity (RDC) in order to monitor crystallinity improvement at a designated 2θ value:

\[
RDC = \frac{Isam}{Iref}
\]

Where Isam is the peak height of the sample under investigation and Iref is the peak height for the reference substance (i.e. plain drug) at the same angle of the highest intensity.

Scanning Electron Microscopy (SEM):

SEM was carried out using JEOL Electron Probe Microanalyzer (JXA-840A, JEOL, Tokyo, Japan) to study the morphological characteristics of the optimized ternary SD and its PM compared to pure DMP. The selected samples were mounted on a double-sided adhesive tape. Gold coating was applied on the surface of particles before examination to render the surface electroconductive.

RESULTS AND DISCUSSION

Phase solubility studies

After UV scanning of DMP in phosphate buffer (pH 6.8), the maximum absorption of DMP in such medium was at wavelength of 284 nm. Figure 2 shows the effect of different carriers (PVP K30, urea and PEG 8000) on the solubility of DMP in phosphate buffer pH 6.8 at 25±0.5°C according to the phase solubility technique.

Determination coefficients (R²) were 0.9875, 0.9969 and 0.9447 for phase solubility diagrams of DMP with PVP K30, urea and PEG 8000, respectively. The solubility of DMP was found to be 10.73 µg/ml and linearly increased as the carrier concentration was increased suggesting the features of an A type solubility phase diagram.

![Figure 2: Phase solubility diagrams of DMP in phosphate buffer pH 6.8 at 25±0.5°C in the presence of increased concentrations of PVP K30, urea and PEG 8000.](image-url)
At 5% w/v of PVP K30, urea and PEG 8000, DMP solubility was increased by 2.20, 1.83 and 1.48 folds, respectively (Table 1). Consequently, these carriers were ranked according to their effect on increasing DMP solubility as PVP K30 > urea > PEG 8000. The increment of drug solubility could be explained by solubilization effect of carriers, their improving influence on drug wettability and through the formation of soluble complexes between hydrophobic drug and hydrophilic carrier\(^a, b\).

**Table 1:** Solubility data of DMP in solutions of different carriers at 25±0.5°C.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>PVP K30</th>
<th>Urea</th>
<th>PEG 8000</th>
</tr>
</thead>
<tbody>
<tr>
<td>Phase solubility diagram type</td>
<td>(A_1)</td>
<td>(A_2)</td>
<td>(A_3)</td>
</tr>
<tr>
<td>Solubility (µg/ml)(^a)</td>
<td>23.64</td>
<td>19.62</td>
<td>15.93</td>
</tr>
<tr>
<td>Solubility factor (^b)</td>
<td>2.20</td>
<td>1.83</td>
<td>1.48</td>
</tr>
</tbody>
</table>

\(^a\) Solubility of DMP in the presence of 5% w/v carrier concentration.
\(^b\) Solubility factor-total solubility of DMP in the presence of 5% w/v carrier concentration/intrinsic solubility of DMP.

Table 2: Drug content uniformity of different DMP systems

<table>
<thead>
<tr>
<th>Formulae</th>
<th>Weight ratio</th>
<th>Drug content %</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>PM(^a)</td>
<td>SD(^e)</td>
</tr>
<tr>
<td>DMP/PEG 8000(^c)</td>
<td>1:1</td>
<td>9.80</td>
</tr>
<tr>
<td></td>
<td>1:5</td>
<td>27.81</td>
</tr>
<tr>
<td></td>
<td>1:9</td>
<td>28.01</td>
</tr>
<tr>
<td>DMP/Urea</td>
<td>1:1</td>
<td>18.61</td>
</tr>
<tr>
<td></td>
<td>1:5</td>
<td>34.62</td>
</tr>
<tr>
<td></td>
<td>1:9</td>
<td>37.02</td>
</tr>
<tr>
<td>DMP/PVP K30(^d)</td>
<td>1:1</td>
<td>30.62</td>
</tr>
<tr>
<td></td>
<td>1:5</td>
<td>53.63</td>
</tr>
<tr>
<td></td>
<td>1:9</td>
<td>52.43</td>
</tr>
<tr>
<td>DMP/PVP K30/PL F-127(^e)</td>
<td>1:9:0.125</td>
<td>32.42</td>
</tr>
<tr>
<td></td>
<td>1:9:0.25</td>
<td>37.42</td>
</tr>
<tr>
<td></td>
<td>1:9:0.5</td>
<td>30.22</td>
</tr>
<tr>
<td>DMP/PVP K30/HPMC E50 LV(^f)</td>
<td>1:9:2.25</td>
<td>23.01</td>
</tr>
<tr>
<td></td>
<td>1:9:4.5</td>
<td>31.82</td>
</tr>
<tr>
<td></td>
<td>1:9:9</td>
<td>33.62</td>
</tr>
<tr>
<td>DMP/PVP K30/PEG 8000</td>
<td>1:9:2.25</td>
<td>28.61</td>
</tr>
<tr>
<td></td>
<td>1:9:4.5</td>
<td>27.81</td>
</tr>
<tr>
<td></td>
<td>1:9:9</td>
<td>27.41</td>
</tr>
</tbody>
</table>

\(^a\) Polyethylene glycol 8000; \(^b\) Polyvinylpyrrolidone K30; \(^c\) Pluronic F-127; \(^d\) Hydroxypropyl methylcellulose E50 LV; \(^e\) Physical mixture and \(^f\) Solid dispersion.

**In-vitro dissolution studies**

Dissolution profiles of the prepared systems are demonstrated in Figures 4-9 and the statistically analyzed PD\(_2\) data are presented in Table 3.

It was evident that the pure drug exhibited a slow dissolution even after 60 minutes where the percentage of drug dissolved after 60 minutes only reached about 6.54±2.66% that could be related to the hydrophobicity, poor wettability and/or agglomeration of DMP particles resulting in floating of drug powder on the surface and consequently hindering its dissolution. On the contrary, PMs as well as SDs immediately sank to the bottom of the dissolution vessels. All carriers had significant effects on PD\(_2\) where the P value was less than 0.05.

As general observations, the dissolution rate of DMP from all PMs was higher than that of the pure drug. The
increased dissolution rate might be attributed to the increased wettability and dispersibility of DMP where the dry mixing brought the drug in close contact with the hydrophilic carrier. Similarly, all SDs showed enhanced dissolution rate compared to pure DMP that might be due to the effect of hydrophilic carriers on drug wettability. Other explanations were related to the solubilization, molecular/colloidal dispersion of drug in the mixture and reduction in the drug crystallinity (i.e. polymorphic transformation of drug crystals) that were obtained via the formulation of solid dispersions.

Table 3: Percentage of drug dissolved after 2 minutes (PD2) in phosphate buffer pH 6.8 of different DMP systems at 37±0.5°C (mean±SD, n=3).

<table>
<thead>
<tr>
<th>System</th>
<th>PM1:1</th>
<th>PM1:5</th>
<th>PM1:9</th>
<th>SD1:1</th>
<th>SD1:5</th>
<th>SD1:9</th>
</tr>
</thead>
<tbody>
<tr>
<td>DMP/PEG 8000</td>
<td>6.20±0.20</td>
<td>17.61±0.20</td>
<td>19.43±0.31</td>
<td>4.47±0.42</td>
<td>10.74±1.72</td>
<td>35.32±0.71</td>
</tr>
<tr>
<td>DMP/PVP K30/PL F-127</td>
<td>1:9:0.125</td>
<td>20.08±0.64</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>DMP/PVP K30/HPMC E50 LV</td>
<td>1:9:0.125</td>
<td>92.71±1.45</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>DMP/PVP K30</td>
<td>1:9:0.25</td>
<td>100.08±1.66</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>DMP/urea</td>
<td>12.27±1.03</td>
<td>25.68±0.31</td>
<td>26.88±0.64</td>
<td>11.00±2.43</td>
<td>35.55±0.31</td>
<td>29.75±1.22</td>
</tr>
<tr>
<td>DMP/PVP K30/PEG 8000</td>
<td>1:9:2.25</td>
<td>8.34±1.29</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>DMP/urea</td>
<td>1:9:4.5</td>
<td>26.95±1.10</td>
<td>17.61±1.11</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>DMP/PVP K30</td>
<td>1:9:0.25</td>
<td>90.51±0.83</td>
<td>75.64±0.72</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>DMP/PVP K30/PEG 8000</td>
<td>1:9:4.5</td>
<td>21.74±0.31</td>
<td>11.14±2.60</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>DMP/urea</td>
<td>1:9:0.25</td>
<td>82.04±3.29</td>
<td>88.64±0.40</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>DMP/Urea</td>
<td>1:9:4.5</td>
<td>88.24±1.83</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

*Domperidone; ‡Polyethylene glycol 8000; §Physical mixture; ¶Solid dispersion; ‖Polyvinylpyrrolidone K30; ‡Pluronic F-127 and ‰Hydroxypropyl methylcellulose E50 LV.

**Binary solid dispersions**

**Domperidone/PEG 8000 systems**

PD2 was significantly enhanced by increasing PEG 8000 concentration in all drug/PEG 8000 systems (p<0.05) till reached the highest value for 1:9 SD where PD2 was 35.32±0.71 (Table 3, Figure 4).

**Domperidone/Urea systems**

As shown in Table 3 and Figure 5, PD2 of drug/urea SDs was significantly increased by increasing urea concentration up to 1:5 weight ratio (p<0.05) where PD2 was 35.55±0.31. After this particular ratio, further increase of urea concentration (i.e. 1:9 SD) resulted in a significant decrement of DMP dissolution rate (p>0.05) where PD2 of 1:9 SD was 29.75±1.22. This might be due to the long time that was consumed by the higher amount of carrier to dissolve.

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**Figure 4:** Dissolution profiles of domperidone from different domperidone/PEG 8000 solid dispersion (SD) and physical mixture (PM) systems in phosphate buffer pH 6.8 at 37±0.5°C.

**Figure 5:** Dissolution profiles of domperidone from different DMP/urea solid dispersion (SD) and physical mixture (PM) systems in phosphate buffer pH 6.8 at 37±0.5°C.
Domperidone/PVP K30 systems:

According to the in-vitro dissolution studies, SD of 1:9 DMP/PVP K30 had the highest significant dissolution rate (p<0.05) compared to other SDs where its PD_2 value was 76.37±0.23 (Table 3, Figure 6). Therefore, this formula was selected to be reformulated as ternary systems using additional water-soluble carriers e.g. PL F-127, HPMC 50 LV and PEG 8000 in different weight ratios by solvent evaporation method.

Domperidone/PVP K30/Pluronic F-127 systems

Ternary systems containing PL F-127 showed significant enhanced dissolution behaviors (p<0.05) by increasing the concentration of PL F-127 reaching maximum PD_2 at weight ratio of 1:9:0.25 DMP/PVP K30/PL F-127 SD (PD_2 was 100.08±1.66) (Table 3 and Figure 7). This might be due to the surfactant property and the great hydrophilicity of PL F-127 resulting in a reduction of the interfacial tension between DMP and dissolution medium, surface availability for rapid dissolution and hence greater wettability of the drug.

Domperidone/PVP K30/HPMC 50 LV systems

Increasing the concentration of HPMC E50 LV up to a certain level resulted in significant enhanced dissolution rate of the drug (p<0.05) (Table 3 and Figure 8). For example, PD_2 values were 26.95±1.10 and 90.51±0.83 for PM and SD of 1:9:4.5 DMP/PVP K30/HPMC E50 LV, respectively.

HPMC is a hydrophilic swellable polymer that is responsible for the formation of highly viscous gelatious barrier diffusion layer at the interface of drug and dissolution medium. Accordingly, further increment of HPMC concentration up to 1:9:9 weight ratio of drug/PVP K30/HPMC E50 LV resulted in a significant decrease in the dissolution rate of PM and SD (p<0.05) where the drug was released slowly from such matrix by diffusion process. For example, PD_2 values were 17.61±1.11 and 75.64±0.72 for PM and SD of 1:9:9 weight ratio, respectively.

Domperidone/PVP K30/PEG 8000 systems

As presented in Table 3 and Figure 9, ternary systems containing PEG 8000 as a second polymer showed a significant increment of PD_2 of DMP up to 1:9:4.5 weight ratio of drug/PVP K30/PEG 8000 (p<0.05). In case of PM, PD_2 of 1:9:9 SD was significantly lower than that of 1:9:4.5 SD (p<0.05). The explanation of this phenomenon might be due to the formation of viscous boundary layer around the drug particles leading to a decrement of DMP dissolution rate. Compared to 1:9:4.5 SD, PD_2 of 1:9:9 SD was decreased with no significant difference between them (p>0.05).
diffusion (Higuchi) models. For example, $R^2$ of 1:5 DMP/urea SD was 0.9556 after being calculated according to Higuchi model. Similarly, $R^2$ values of 1:9 DMP/PVP K30 and 1:9:0.25 DMP/PVP K30/PL F-127 solid dispersions were in accordance with Higuchi model where they were 0.9848 and 0.9523, respectively.

Fourier-transform infrared spectroscopy (FTIR)

In order to get indication on the feasible interaction of the drug with the studied PVP K30 and PL F-127, FTIR analysis was employed (Figure 10). The FTIR spectrum of plain DMP was characterized by N-H stretching at (3119.3 cm$^{-1}$), asymmetric C-H stretching at (2939.95 cm$^{-1}$), symmetric C-H stretching at (2820.38 cm$^{-1}$), N-H deformation at (1697.05 cm$^{-1}$), aromatic C-H stretching at (3022.87 cm$^{-1}$), C=C at (1622.02 cm$^{-1}$) and N=C stretching peak at (1485.88 cm$^{-1}$). The spectrum of PVP K30 showed C-H stretching band at (2953 cm$^{-1}$), C=O band at (1666.20 cm$^{-1}$) and a very broad endothermic band at (3048-3750 cm$^{-1}$) that was related to the presence of water confirming the broad endotherm detected later in DSC study. FTIR spectrum of PL F-127 is characterized by principal absorption peaks of aliphatic C-H stretching at (2886.92 cm$^{-1}$), in-plane O-H bend at (1355.71 cm$^{-1}$) and C-O stretching at (1110.8 cm$^{-1}$).

The FTIR spectra of the optimized ternary SD and PM showed the disappearance of N-H stretching peak of DMP with slight shifting of PVP carbonyl band from (1666.20 cm$^{-1}$) to (1664.27 cm$^{-1}$) and (1662.34 cm$^{-1}$) for PM and SD, respectively. This might indicate an intermolecular hydrogen bonding between =NH group of DMP and the C=O band of PVP in the drug-polymer systems$^{45,46}$.

Differential scanning calorimetry (DSC)

As shown in Figure 11, DSC thermogram of DMP presents a sharp endothermic peak at 243.43°C corresponding to the melting point of the drug. A broad endothermic peak corresponding to PVP K30 was observed at 80.15°C that might be attributed to the loss of water from the
Hygroscopic PVP K30. Pluronic F-127 has an endothermic peak at 57.39°C related to its melting point.

The DSC thermograms of SD and PM showed a disappearance of the drug peak. The absence of DMP endotherm in PM suggested the dissolution of the crystalline drug particles within the molten polymer due to the heating phase during analysis. In case of SD, the absence of DMP endotherm might be due to the formation of solid dispersion of the drug in the presence of water-soluble polymer where the drug could be transformed into an amorphous state. This amorphousness might be related to the intermolecular hydrogen bonding between DMP and PVP K30 and/or loss of drug mobility where the drug was entrapped in polymer after evaporation of solvent.30,47

Figure 11: DSC thermograms of (A) Pure domperidone (DMP), (B) Polyvinyl pyrrolidone K30 (PVP K30), (C) Pluronic F-127 (PL F-127), (D) Physical mixture of 1:9:0.25 DMP/PVP K30/PL F-127 and (E) Solid dispersion of 1:9:0.25 DMP/PVP K30/PL F-127.

Powder X-ray diffraction (PXRD):

Figure 12 shows the PXRD patterns of DMP solid systems. The diffraction spectrum of pure DMP shows its crystalline nature that was demonstrated by numerous sharp, highly intense and less diffused peaks. These peaks were observed at 2θ values of 9.22°, 11.77°, 13.90°, 14.88°, 15.53°, 19.00°, 19.75°, 22.58°, 24.76°, 28.98°, 31.47° and 42.61° in fingerprint regions referring to its crystallinity. A hollow pattern with no diffraction peaks was recorded for PVP K30 indicating its amorphous state. The diffraction spectrum of PL F-127 shows two characteristic peaks at 2θ values of 19.07° and 23.24° indicating the crystalline nature of PL F-127. The position of characteristic peaks of the crystalline polymer was not changed in PM and SD suggesting no change of its polymorph. PXRD patterns of ternary PM and SD exhibited 'halo' shaped diffractograms characterizing the amorphous material since the reflexes did not return to the base line. Furthermore, broadening of DMP peaks and reduction of their intensities were observed suggesting the conversion of crystalline DMP to partially disordered molecules.30

Figure 12: PXRD patterns of (A) Pure domperidone (DMP), (B) Polyvinyl pyrrolidone K30 (PVP K30), (C) Pluronic F-127 (PL F-127), (D) Physical mixture of 1:9:0.25 DMP/PVP K30/PL F-127 and (E) Solid dispersion of 1:9:0.25 DMP/PVP K30/PL F-127.

Peak height of DMP at 22.6° 2θ was selected to calculate the RDC of DMP, best ternary PM and ternary SD. When pure DMP was considered as a reference sample, a significant decrement in crystallinity of the characterized ternary systems was observed (p<0.05). RDC values were 1, 0.17 and 0.14 for pure DMP, ternary PM and ternary SD, respectively indicating the amorphousness of DMP and the formation of SD as previously investigated by PXRD patterns.

Table 5: Relative degree of crystallinity (RDC) values of domperidone/polyvinylpyrrolidone K30/Pluronic F-127 systems at a degree of 2θ= 22.6°.

<table>
<thead>
<tr>
<th>Formula</th>
<th>RDC(^2) at 2θ=22.6°</th>
</tr>
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<tbody>
<tr>
<td>DMP(^b)</td>
<td>1</td>
</tr>
<tr>
<td>PM(^c)</td>
<td>0.17</td>
</tr>
<tr>
<td>DMP/PVP K30(^d)/PL F-127(^e)</td>
<td>0.14</td>
</tr>
</tbody>
</table>

\(^{a}\) Relative degree of crystallinity; \(^{b}\) Domperidone; \(^{c}\) Physical mixture; \(^{d}\) Polyvinylpyrrolidone K30; \(^{e}\) Pluronic F-127 and \(^{f}\) Solid dispersion.

Scanning electron microscopy (SEM)

SEM micrographs that reveal the surface morphology of scanned samples at 1000X are shown in Figure 13. SEM micrograph of pure DMP shows crystalline particles of rather irregular shape and size (Figure 13A), while the SEM micrograph of PM reveals more identified cotton-shaped powder with crystalline dusts of DMP deposit on the surface (Figure 13B). SD appeared in the form of irregular particles in which the original crystalline morphology of DMP disappeared and small lumps of amorphous pieces of irregular size were present (Figure 13C). This result could be attributed to dispersion of the drug in the polymer matrix confirming the findings based on PXRD patterns. The change in structure might be one of the causes for the increased dissolution rate.
plexes of water

Suir ML, Marchetti


CONCLUSION

This study demonstrated the possibility of improving DMP solubility and dissolution performance by the formulation of solid dispersions. The binary solid dispersion of 1:9 DMP/PVP K30 achieved the highest significant percentage of drug dissolved after 2 minutes compared to all binary systems. This weight ratio was selected to formulate ternary solid systems by incorporating other water-soluble carriers. Ternary SD of 1:9:0.25 DMP/PVP K30 achieved approximately 100% drug dissolved over the first 2 minutes. Treatment of dissolution data according to zero, first order and Higuchi model resulted in determination coefficient values subjected to diffusion release kinetics. In-vitro dissolution studies, FTIR, DSC, PXRD and SEM analysis revealed the amorphization of DMP and the formation of intermolecular hydrogen bond between the drug and PVP K30 that might be responsible for dissolution enhancement.

REFERENCES


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